

# CDP-CHOLINE NEUROPROTECTION AND VASCULAR REMODELLING VIA IRS-1 MECHANISMS IN VASCULAR DEMENTIA

## GROWING NEURONS ATOP MICROELECTRONIC CHIPS AS A NEW MODEL FOR NEURODEGENERATION

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### INTRODUCTION

Evidence of reduced blood-brain barrier (BBB) integrity preceding other Alzheimer's disease (AD) pathology provides a strong link between cerebrovascular pathology and AD. In animals models, amyloid- $\beta$  peptide - injected animals exhibited a commonality in perturbations of microvessels compared with those evident in AD brain<sup>(22)</sup>. It was suggested that amyloidogenesis promotes extensive neoangiogenesis leading to increased vascular permeability and subsequent hypervascularization in AD. In human patients hypervascularity was corroborated in a comparison of postmortem brain tissues from AD. Brain microvessels derived from patients with AD expressed numerous factors implicated in vascular activation and angiogenesis. Signaling cascades associated with vascular activation and angiogenesis were upregulated in AD-derived brain microvessels<sup>(21)</sup>. However, these newly formed blood vessels may be non-functional. All above provides a new paradigm for integrating vascular remodeling with the pathophysiology observed in AD<sup>(20)</sup>. Therefore, vascular activation hypothesis could be a novel, unexplored therapeutic target in AD.

**Background of our own recent investigations:** In our recent study, we have, for the first time, demonstrated both a vascular protective, and proangiogenic effect of citicoline using *in vivo* and *in vitro* models<sup>(19)</sup>. Our data suggests a strong protective effect against the damaging process of excitotoxicity and hypoxia, similar to that experienced after acute ischaemic stroke. In regard to the possible mechanism our protein studies demonstrated that citicoline induced pERK1/2 expression, a key mitogenic signalling protein known to be involved in angiogenesis and generally stimulated by growth factors through interaction with their receptors<sup>(23)</sup>.

There is the potential of citicoline to activate intra-cellular signal transduction pathways and induce phosphorylation of down-stream angiogenic molecules; hence we investigated this ability in more detail by analysis of the Kinexus-phospho-protein Western screening following treatment of vascular EC with citicoline.

Interestingly, treatment with citicoline modified the expression of only several of the >500 proteins on the array showing a degree of specificity. Insulin receptor substrate-1 (IRS-1) was phosphorylated in

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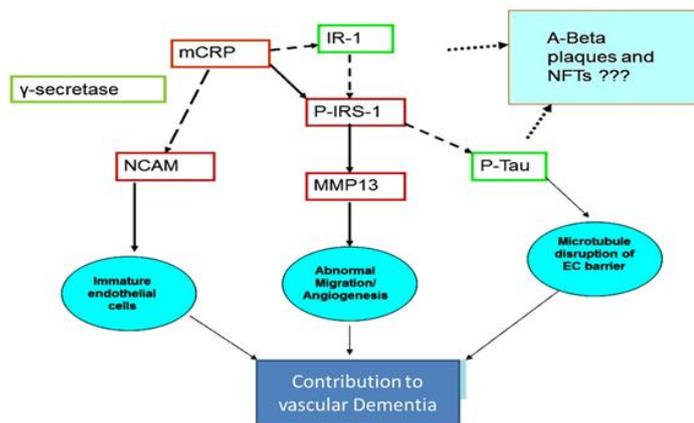
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the presence of citicoline. IRS-1 over-expression was attributed to increased angiogenesis in human EC in association with increased Akt and VEGF-A expression [Stephens et al, 2012], whilst *in vivo*, antisense IRS-1 sequences delivered by sub-conjunctival injection inhibited rat corneal neovascularisation<sup>(18)</sup>, and when delivered by means of eye-drops (GS-101) were found to be tolerable in a phase-1 clinical trial and may be sufficient to prevent neovascularisation in disease such as retinopathy and neovascular glaucoma<sup>(15)</sup>. Therefore, IRS-1 represents a potent modulator of pro-angiogenic signalling cascades in vascular EC and as such, since we have shown both *in vitro*, and in the rat

chronic re-vascularization after stroke impacting indirectly but significantly also on neuronal survival and re-integration<sup>(17)</sup>. Figure one for details<sup>(19)</sup>.

**IRS-1 in neurodegeneration:** Only recently, it has been demonstrated that beta-amyloid (Aβ) oligomers are implicated in Alzheimer's disease leading to phosphorylation and degradation of the adaptor protein insulin receptor substrate-1 (IRS-1). IRS-1 couples insulin and other trophic factor receptors to downstream kinases and neuroprotective signaling. Increased phospho-IRS-1 is found in AD brain. Levels of IRS-1 and their activated kinases correlated positively with those of

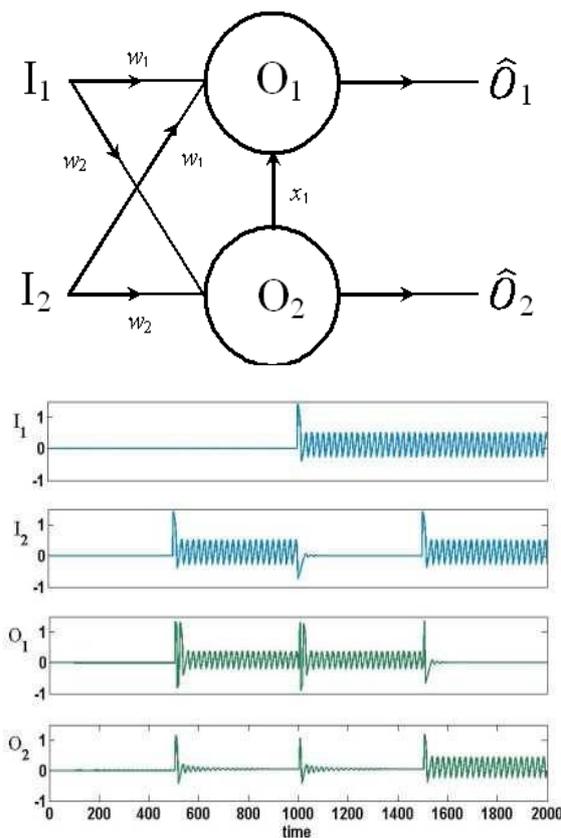


**Figure 1 shows the operation of a binary half-adder based on Fitzhugh-Nagumo oscillators (which are simplified versions of the Hodgkin-Huxley models).**

model of temporary MCAO that citicoline induces phosphorylation of IRS-1 and concomitant EC activation and increased vascularisation. This could be a key novel mechanism of action of citicoline implicated in stroke recovery pathways and angiogenesis until now. This may be an extremely valuable novel finding in regard to understanding the potential mechanisms through which citicoline treatment results in patient recovery, since both protection of EC and induction and maintenance of angiogenesis is key to both short-term and

oligomeric Aβ plaques and are negatively associated with episodic and working memory, even after adjusting for Aβ plaques, neurofibrillary tangles, and APOE ε4. Brain insulin resistance thus appears to be an early and common feature of AD, a phenomenon accompanied by IGF-1 resistance and closely associated with IRS-1 dysfunction potentially triggered by Aβ oligomers and yet promoting cognitive decline independent of classic AD pathology<sup>(17,18)</sup>

**New assays for neuronal degeneration:** It is now understood that periodic behaviour is not confined to a limited number of physiological processes but is abundant in most biological systems. Periodicity in processes of the human body encompass phenomena such as genetic interactions, heartbeat rhythms, oscillating secretory, retina and muscle cells, cytoskeletal structures, bacterial oscillations, rhythmic



**Figure 2:** (a) Schematic of a binary oscillator half-adder. (b) Numerical simulation: Time series showing correct logic function of a half-adder. An oscillation corresponds to a 1 in binary and no oscillation is a zero. The sum oscillator  $O_1$  will oscillate if either  $I_1$  or  $I_2$  is active. The carry oscillator  $O_2$  will oscillate if both  $I_1$  and  $I_2$  are active. An inhibitory connection, say  $x_1$ , from  $O_2$  to  $O_1$  suppresses oscillator  $O_1$  if  $O_2$  is active.

behaviour in growth and development, and most importantly for this study, neuronal

oscillations<sup>(1-3)</sup>. In 1952, whilst modelling neurons, Hodgkin and Huxley were able to accurately model the action potential in the giant squid axon<sup>(4)</sup>. Their nonlinear ordinary differential equations approximate electrical characteristics of excitable oscillatory cells such as cardiomyocytes and neurons. In 2009, Borresen and Lynch<sup>5</sup> published a paper suggesting a novel idea based on biological neural computing utilising the Hodgkin-Huxley equations, and following this work, three years later UK and International patents were published<sup>(6)</sup>. The patent illustrates how it is possible to construct binary logic gates from biological neurons grown on a chip. It is proposed that neurons could be grown on a chip and that they could be trained to perform certain logical operations. Binary oscillator logic gates could then be used to test drugs that may halt or reverse symptoms of neurological disorders such as Alzheimer's, Parkinson's disease and epilepsy. A number of research groups are now able to grow neurons on chips using a variety of techniques including an aligned micro-contact printing technique, patch clamping (which yields very accurate information but is invasive) and extracellular recordings by means of external micro-transducers or optical measurements (which are non-invasive)<sup>(8-13)</sup>, and it has recently been shown that Parkin diseased neurons can also be grown on a chip<sup>(14)</sup>. In 2012<sup>(15)</sup>, it was shown that citicoline could be used as a protective treatment against Alzheimer's following a stroke.

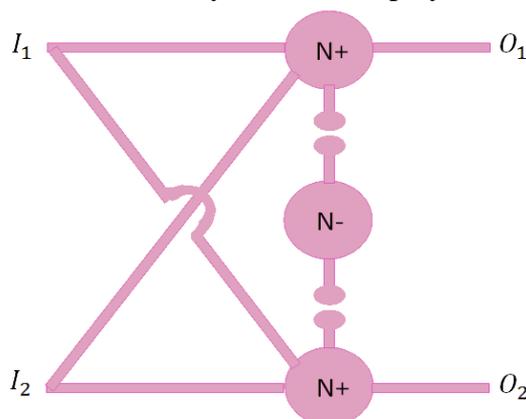
Based on our previous results and citicoline involvement in blood vessel remodelling via IRS-1 pathway together with a new evidence of vascular hypothesis in AD, we aim to develop an *in vitro* assay for neuronal degradation of Alzheimer's diseased neurons. In this model we will study effects of citicoline/IRS-1 pathways on neurones grown on atop microelectronic chips in conditions of ischemia and

neurodegeneration. It is a multidisciplinary project drawing on Medicine, Biology, Mathematics, Chemistry, Physics, Engineering and Computing.

### FUTURE DEVELOPMENTS

We have already shown that it is possible to construct logic gates and memory using threshold oscillator logic<sup>(16)</sup>. We demonstrated how coupled threshold oscillators (neurons) may be used to perform binary logic in a manner entirely consistent with modern computer architectures.

Figure 2(a) demonstrates a viable circuit schematic for half-adder implementation using two neuronal oscillators labelled  $O_1$  and  $O_2$  and two inputs  $I_1$  and  $I_2$ , which may themselves be the output from other neurons in a more complex circuit. In order to perform the logical operations it is necessary that either neurons with differing thresholds be used or the connections between the neurons should be of differing weights, indicated by  $w_1$  and  $w_2$  in the figure. Figure 2(b) displays a time series plot of a binary half-adder. Schematics and time series for a two oscillator full adder, a three oscillator seven input full adder, a 2x2 bit binary multiplier and a set reset flip-flop (used for memory) are also displayed<sup>(16)</sup>.



**Figure 3:** Schematic of a binary oscillator half-adder using biological neurons. Neurons N+ depict excitatory neurons and neuron N- depicts an inhibitory interneuron.

Using the logic circuitry highlighted by Lynch et al<sup>(6,7,16)</sup>, we believe it is possible to build an assay for neuronal degradation. Implementation of the half-adder using biological neurons on a chip will require three neurons as neurons are either excitatory or inhibitory, they cannot be both. Figure 3 shows a schematic of the biological half-adder.

### CONCLUSIONS

**Future work:** Our current work is structured in five main steps:

1. Growing neurons on chips.
2. Design of a new model of threshold logic based on biological neurons.
3. Simulation of threshold oscillator logic using a suitable mathematical package.
4. Building neuronal logic circuits using neurons.
5. Growth of Alzheimer/vascular dementia diseased neurons on chip and measurement of neuronal degradation for different citicoline dosages.

This methodology will allow for the sub-cellular resolution (micro-pixel) within neurobiological preparations for example neuron-neuron interfaces and indeed later, entire neuronal networks. This proposed methodology has potential application in all areas of neuroscience, medical diagnostics and pharmacology.

### REFERENCES

1. Rapp P.E. An atlas of cellular oscillators, J. Exp. Biology 1979; 81: 281-306.
2. Hierlemann A., Frey U., Hafizovic S., Heer F. Growing cells atop microelectronic chips: Interfacing electrogenic cells in vitro with CMOS-based microelectrode arrays introduction, Proc. of IEEE 2011; 99: 249-251.
3. Kruse K. and Jülicher F. Oscillations in cell biology, Opinion in Cell Biology 2005; 17: 20–26.
4. Hodgkin A. and Huxley A. A quantitative description of membrane current and its

- application to conduction and excitation in nerve, *J. Physiol.* 1952; 117: 500-544.
5. Borresen J. and Lynch S. Neuronal computers, *Nonlinear Anal. Theory, Meth. & Appl.* 2009; 71:2372-2376.
  6. Lynch S. and Borresen J. Binary half adder using oscillators, International Publication Number WO 2012/001372 A1, 1-57.
  7. Lynch S. and Borresen J. Binary half-adder and other logic circuits, UK Patent Number 2012; GB 2481717 A, 1-57.
  8. James CD, Davis R., Meyer M. Aligned microcontact printing of micrometer-scale poly-L-lysine structures for controlled growth of cultured neurons on planar microelectrode arrays, *IEEE Trans. on Biomedical Engineering* 2000; 47: 17-21.
  9. Delivopoulos E, Murray AF, MacLeod N.K. and Curtis JC. Guided growth of neurons and glia using microfabricated patterns of parylene-C on a silicon dioxide background, *Biomaterials* 2009; 30: 2048-2058.
  10. Xu F.L., Fakas S., Korbeek S. et al. Mercury-induced toxicity of rat cortical neurons is mediated through N-methyl-D-Aspartate receptors, *Molecular Brain* (2012); 5: 1-14.
  11. Fromherz P., Eick S. and Hofmann B. Neuroelectronic Interfacing with Semiconductor Chips, *Nanoelectronics and Information Technology*, Ed. R Waser, 2<sup>nd</sup> Edition. 2012.
  12. Stephens C.L., Toda H., Palmer T.D. Adult neural progenitor cells reactivate superbursting in mature neural networks, *Experimental Neurology* 2012; 234: 20-30.
  13. Arthur J.V., Merolla P.A., Akopyan F. Building block of a programmable neuromorphic substrate: A digital neurosynaptic core, International Joint Conference on Neural Networks 2012.
  14. Jiang HB., Ren Y., Yuen EY. Parkin controls dopamine utilization in human midbrain dopaminergic neurons derived from induced pluripotent stem cells, *Nature Communications* 2012; 3: Article number 668.
  15. Krupinski J., Abudawood M., Matou-Nasri S. Citicoline induces angiogenesis improving survival of vascular/human brain microvessel endothelial cells through pathways involving ERK1/2 and insulin receptor substrate-1, *Vascular Cell* 2012; 4: 20.
  16. Borresen J. and Lynch S. Oscillatory threshold logic, *PLoS ONE* 2012; 7: e48498.
  17. Talbot K, Wang HY, Kazi H, Han LY, Bakshi KP, Stucky A, Fuino RL, Kawaguchi KR, Samoyedny AJ, Wilson RS, Arvanitakis Z, Schneider JA, Wolf BA, Bennett DA, Trojanowski JQ, Arnold SE. Demonstrated brain insulin resistance in Alzheimer's disease patients is associated with IGF-1 resistance, IRS-1 dysregulation, and cognitive decline. *J Clin Invest.* 2012 Apr 2;122(4):1316-38. doi: 10.1172/JCI59903.
  18. O'Neill C, Kiely AP, Coakley MF, Manning S, Long-Smith CM Insulin and IGF-1 signalling: longevity, protein homeostasis and Alzheimer's disease. *Biochem Soc Trans.* 2012 Aug ;40(4):721-7. doi: 10.1042/BST20120080.
  19. Krupinski J, Abudawood M, Matou-Nasri S, Al-Baradie R, Petcu E, Justicia C, Planas A, Liu D, Rovira N, Grau-Slevin M, Secades J, Slevin M. Citicoline induces angiogenesis improving survival of vascular/human brain microvessel endothelial cells through pathways involving ERK1/2 and insulin receptor substrate-1. *Vasc Cell.* 2012 Dec 10;4(1):20. [Epub ahead of print]
  20. Biron KE, Dickstein DL, Gopaul R, Jefferies W. *PLoS One.* 2011;6(8):e23789. doi: 10.1371/journal.pone.0023789. Epub 2011 Aug 31. Amyloid triggers extensive cerebral angiogenesis causing blood brain barrier permeability and hypervascularity in Alzheimer's disease.
  21. Grammas P, Sanchez A, Tripathy D, Luo E, Martinez J. Vascular signaling abnormalities in Alzheimer disease. *Cleve Clin J Med.* 2011 Aug;78 Suppl 1:S50-3. doi: 10.3949/ccjm.78.s1.09.
  22. Jantaratnotai N, Ryu JK, Schwab C, McGeer PL, McLarnon JG. Comparison of Vascular Perturbations in an A $\beta$ -Injected Animal Model and in AD Brain. *Int J Alzheimers Dis.* 2011;2011:918280. doi: 10.4061/2011/918280. Epub 2011 Sep 29.
  23. Ma QL, Yang F, Rosario ER, Ubada OJ, Beech W, Gant DJ, Chen PP, Hudspeth B, Chen C, Zhao Y, Vinters HV, Frautschy



SA, Cole GM. Beta-amyloid oligomers induce phosphorylation of tau and inactivation of insulin receptor substrate via c-Jun N-terminal kinase signaling:

suppression by omega-3 fatty acids and curcumin. *J Neurosci.* 2009 Jul 15;29(28):9078-89. doi: 10.1523/JNEUROSCI.1071-09.2009.